THE GERMACRANOLIDES OF VIGUIERA BUDDLEIAEFORMIS STRUCTURES OF BUDLEIN-A AND -B[‡]

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Abstract—The structure of budlein-A, the main sesquiterpene lactone of Viguiera buddleiaeformis was established as the 8 angeloyl ester of 1 keto, 8- β , 14-dihydroxy germacra-2,4,11 (13)-trien-3, (10 β) oxido-6 α , 12-olide. Its structure and stereochemistry was determined by chemical and spectroscopic means. Budlein-B, found in the same plant as a minor constituent, is 8 α , 15-dihydroxygermacra-1 (10), 4, 11 (13)-trien-6 α , 12-olide.

In a previous paper we described two germacranolides, viguestenin and desacetylviguestenin, isolated from Viguiera stenoloba [1]. The present article deals with the isolation and structure determination of the 3(2H) furanone germacranolide budlein-A 1a, a constituent of V. buddleiaeformis. Budlein-B was also isolated in low yield from the same plant; its structure and stereochemistry was found to be as depicted in 6a.

Budlein-A (1a) $C_{20}\bar{H}_{22}O_7$, mp 106–108°; $[\alpha]_D - 82.3^\circ$ MeOH, showed in the IR spectrum the characteristic bands of a 3(2H) furanone moiety [2,3] (1710 cm⁻¹ keto group and 1590 cm⁻¹ enolic double bond). The UV absorption (λ_{max} 266 nm, ϵ , 10000) indicated the conjugation of an additional double bond at C-4 with the 3(2H) furanone system, similar to that observed with calaxin and ciliarin [2] (zexbrevin which lacks the C-4 double bond absorbs at 259 nm) [3]. Budlein-A exhibited IR bands at 1760, 1650, 890 cm⁻¹ and a pair of low field doublets in the PMR spectrum at 5.70 ppm (1H, J 2 Hz) and 6.30 ppm (1 H, J 2 Hz), characteristic of a y-lactone conjugated with an exocyclic methylene group (UV absorption λ_{max} 215 nm, ϵ , 20800 both, lactone and ester chromophore). The presence of a OH group was indicated by an IR band at 3400 cm⁻¹ and the formation of an acetate. In the MS, compound 1a showed a M+ at m/e 374 and other prominent peaks at m/e 274 (M^+-100) , m/e 55 and a base peak at m/e 83 (Me-CH=C(Me)C≡O). These fragmentations are typical of the cleavage of esters of angelic, tiglic and senecioic acids. In the PMR spectrum of budlein-A a multiplet appeared at 6.08 ppm (1 H), a chemical shift typical of the vinyl proton of angelic acid. The vinyl hydrogen of tiglic acid appears at 7.0 ppm whereas the α-vinyl proton of senecioic acid resonates at higher field [4].

The PMR spectrum of budlein-A showed only signals for the C-10 Me group and the vinyl Me groups of the

The three previously described 3(2 H) furanone germacranolides, calaxin, ciliarin [2] and zexbrevin [3], contain a C-6 lactone closure, thus making structure 1a more probable. This was confirmed in the following manner: in the PMR spectrum of compound 1a, the signal at 5.33 ppm (1 H) was attributed to the C-6 allylic hydrogen of the lactone closure, since it was shifted upfield to 47 ppm (the typical position for the lactonic proton) in tetrahydrobudlein-A 2. The multiplet centered at 5.23 ppm (1 H) in 1a remained unchanged in compound 2 and was ascribed to the hydrogen at the carbon bearing the ester group. When the signal at 5.23 ppm was irradiated, the multiplet at 3.75 (proton at C-7) was simplified; at the same time the ABX system (C-9 methylene) collapsed to an AB system. This demonstrated that the signal at 5.23 corresponds to C-8 proton (ester base), the usual position for the ester group in lactones of this type [2,3,5]; therefore budlein-A is represented by 1a.

Catalytic hydrogenation of 1a was very complicated due to the presence of four double bonds and a hydrogenolizable allylic alcohol. After several attempts, there was obtained in a very low yield the tetrahydroderivative 2. Compound 2 still contains the 3(2 H) furanone moiety (IR bands at 1710 and 1590 cm⁻¹) and the ester double bond (m/e 83). The saturation of the exocyclic methylene

angelate ester moiety (Table 1). The typical signal for the C-4 Me group, common for lactones of this type [2], was not present; instead, the spectrum exhibited a broad singlet at 4:36 ppm (2 H) which was assigned to the -CH₂-O- protons of an allylic primary alcohol. The vinyl hydrogen at C-5 appeared as a multiplet centered at 6·17 ppm (1 H, $J_{5.6}$ 4 and $J_{5.14}$ 1·5 Hz); the oxygen bridge of the dihydrofuranone system must lie between C-3 and C-10 of the germacranolide skeleton, since the C-10 Me group appeared as a singlet at 1.48 ppm (3H). Two overlapping complex signals centered at 5.33 and 5.23 ppm (1 H each) were ascribed to C-6 and C-8 protons. From the foregoing discussion we can assign structure 1a for budlein-A, although at this point the alternative structure with the ester group attached to C-6 and the lactone closed to C-8 cannot be excluded.

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group was indicated by the absence of the low field vinyl proton doublets.

(6b) R = Ac

Sodium borohydride reduction of 1a afforded octahydrobudlein-A 3 in which the carbonyl group at C-1 and the C-4 and C-11 double bonds were reduced. The ester double bond remained intact as indicated by the complex signal centered at 6·0 ppm (1 H). The C-6 hydrogen in 3 appeared as a multiplet centered at 4·5 ppm (1 H); the signal corresponding to the C-8 proton remained unchanged at 5·23 ppm.

Acetylation of budlein-A in the usual manner gave an unseparable mixture of several compounds. Budlein-A acetate 1b was obtained when the acetylation was carried out under more basic conditions using triethylamine. The PMR spectrum of 1b showed a singlet at 2·15 ppm (3 H) indicative of the acetoxy group; a singlet at 4·85 ppm (2 H) was assigned to the $-CH_2-O-$ protons at the base of the acetate. The MS showed a base peak at m/e 83 and other prominent peaks at m/e 356 (M⁺-AcOH) and m/e 43 (Me-C \equiv O⁺).

Catalytic hydrogenation of 1b over palladium-charcoal gave a mixture of 3 compounds. Separation by TLC afforded in low yield hexahydrobudlein-A acetate 4. In the vinyl proton region, 4 showed only the signal corresponding to the C-2 proton, thus indicating a complete saturation of the remaining double bonds.

Treatment of budlein-A with m-chloroperbenzoic acid afforded the epoxide 5, in which only the ester double bond was epoxidized. In the PMR spectrum, budlein-A epoxide exhibited a doublet at 1·3 ppm (3 H, J 5 Hz), a singlet at 1·2 ppm (3 H) and a quartet centered at 3·1 ppm (1 H, J 4 and 5 Hz) attributed to the Me groups and the proton of the epoxide, respectively.

The stereochemistry of budlein-A at C-6 and C-8 was established from the dihedral angles (Table 1) calculated from the coupling constants. In the PMR spectra of compounds 1a, 1b, 2 and 3, the signal corresponding to the C-7 proton has an abnormally low chemical shift (3.72

Table 1. PMR spectrum of budlein-A

Proton number	Chemical shift	Multiplicity	Coupling constants	Dihedral angles
H-2	5.65	s		
H-5	6.17	t, d	$J_{5,14} = 1.5$	
H-6	5 33	m	$J_{6.7} = 4.5$	47° or 118°
			$J_{5,6} = 40$	50° or 120°
H-7	3.75	m	$J_{7,13}=2.8$	
			$J_{7,13} = 30$	
H-8	5 23	m	$J_{8,9} = 5.5$	42° or 125°
			$ \begin{vmatrix} J_{8,9} &= 40 \\ J_{7,8} &= 20 \end{vmatrix} $	<u>50°</u> or 120°
				63° or <u>110°</u>
H-9	2.51	d, d	$J_{9,9} = 15$	
H-9'	2.32	d, d	1	
14CH ₂ -O	4 36	d, t		
15CH ₃	1.48	S		
H-18	6 08	q, q	$J_{16,17} = 1.5$	
			$J_{16,18} = 7$	
19CH ₃	1-91	m		
20CH ₃	1.78	m		

Spectra were determined in CDCl₃ at 100 MHz, Values are given in ppm (δ scale) relative to TMS as an internal standard. Signals are described as follows: s = singlet, d = doublet; t = triplet, q = quartet, m = multiplet. Coupling constants are given in Hz.

ppm), indicating that this hydrogen is close to the oxygen of the furan ring. If the usual assumption is made that the C-7 side chain is β -oriented, as in all sesquiterpene lactones of known stereochemistry, this proximity requires an α -oriented C-7 hydrogen, an α C-10 methyl group and a cis C-4 double bond (construction of a Dreiding model containing a trans C-4 double bond proved impossible). The calculated dihedral angle of 118° at the C₆-C₇ bond (the same angle was observed for zexbrevin) [3] indicated a β -configuration for the C-6 hydrogen or a trans lactone closure.

The configuration of the oxygen function at C-8 was found to be β -oriented, since the coupling constants between H₇-H₈, H₈-H₉ and H₈-H₉, have small values. This is in agreement with the required dihedral angles (Table 1). Hence the structure and the stereochemistry for budlein-A must be as depicted in 1a.

Budlein-B (6a) $C_{15}H_{20}O_4$, mp 162° ; $[\alpha]_D + 3.1^\circ$ MeOH, was isolated from the more polar fractions. It contains a y-lactone conjugated with an exocyclic methylene group, as indicated by the IR bands (1765, 1665, 890 cm⁻¹) and the characteristic pair of low field doubles in the PMR spectrum at 5.60 ppm (1 H, J 1.5 Hz) and 6.30 ppm (1 H, J 2 Hz). The UV spectrum (λ_{max} 222 nm, ε , 8908) was in accord with the presence of an α , β -unsaturated y-lactone. In the MS, compound 6a showed a M^+ at m/e 264 and another prominent peak at m/e 246 (M^+-H_2O) and m/e 231 $(M^+-18-15)$. The presence of two OH groups was indicated by the strong IR band at 3400 cm⁻¹ and the formation of a diacetate. The PMR spectrum exhibited a doublet centered at 1.68 ppm (3 H, J 1 Hz), ascribed to a vinyl Me group. The six allylic protons appeared as a five proton multiplet centered at 2.28 ppm (C-2, 2 H; C-3, 2 H; C-9, 1 H); the complex signal at 2.82 ppm (2 H) was assigned to the C-7 and C-9 protons. In the low field region of the spectrum the vinyl protons at C-1 and C-5 appeared as complex signals at 5:15 ppm and 4:85 ppm, respectively. The expected signal for the vinyl Me group at C-10, found in the majority of the germacradienolides [5] was not present; instead, the spectrum exhibited a sharp symmetrical AB quartet at 3.9 ppm (2 H, J 1 and 5 Hz), assigned to the OH-methylene group. The broad quartet

at 50 ppm (1 H) was attributed to the C-6 lactonic proton.

Budlein-B, upon acetylation with pyridine-acetic anhydride, afforded budlein-B diacetate 6b which was identical in all respects with zexbrevin D, previously isolated from Zexmenia brevifolia [6].

The stereochemistry at C-8 of budlein-B was established by the method developed by Horeau for determining the configuration of asymmetric centers containing a secondary OH group. Treatment of budlein-B with excess racemic α -phenylbutyric anhydride and work up in the usual manner afforded a laevorotatory optically active acid. Based on the Horeau procedure [7], the asymmetric center at C-8 must have the absolute configuration S (α -oriented OH group).

From all evidence presented above, we propose structure 6a for budlein-B.

EXPERIMENTAL

Mp's are uncorrected. IR spectra were recorded in CHCl₃ and UV in 95% EtOH unless otherwise stated. Analyses were determined by Dr. Alfred Bernhardt, Max Planck Institute fuer Kohlenforschung, Muelheim, West Germany.

Isolation of budlein-A (1a) and budlein-B (6a). Viguiera buddleiaeformis (D.C.) Benth et Hook (10 kg) dried and ground was extracted 2× with EtOH and worked up in the usual manner [3], leaving an oily residue which was dissolved in hot C_6H_6 and chromatographed over Alcoa F-20 alumina. The fractions eluted with CHCl₃-Me₂CO (9:1) were combined and evaporated to dryness giving an amorphous residue; recrystallization from Me₂CO-isopropyl ether afforded pure budlein-A (3·65 g); mp 106-108°; $\lceil \alpha \rceil_0^{15} \cdot 82\cdot3^\circ$ (MeOH); λ_{max} 215, 266 nm; ϵ , 20800 and 10000; ν_{max} 3430, 1770, 1710, 1660, 1590 and 890 cm⁻¹. (Found: C, 64·03; H, 5·98; O, 29·99. $C_{20}H_{22}O_7$ requires: C, 64·15; H, 5·88; O, 29·94%). From the fractions eluted with CHCl₃-Me₂CO (4:1) there was obtained 400 mg of budlein-B; mp 162°; $\lceil \alpha \rceil_0^{15} + 3\cdot1^\circ$ (MeOH); λ_{max} 224 nm; ϵ , 7,200; ν_{max} 3350, 3450, 1765, 1665, 1150 and 1050 cm⁻¹. (Found: C, 68·30; H, 7·51; O, 24·32. $C_{15}H_{20}O_4$ requires: C, 68·28; H, 7·55; O, 24·27%).

Bullein-A acetate (1b). To a soln of budlein-A (175 mg) in CH₂Cl₂ (2·3 ml) was added Et₃N (0·58 ml) Ac₂O (0·23 ml) and C₅H₅N (0·015 ml). The mixture was agitated for 1·5 hr at room temp. Work up in the usual manner afforded a mixture of two compounds: 1b was obtained by preparative-TLC, yield 130 mg; mp 114-116°; [α] $_{6}^{2.5}$ - 78·8° (MeOH); ν _{max} 1740 cm⁻¹ (acetate); M⁺ 412. (Found: C, 63·37; H, 5·88; O, 30·69. C₂₂H₂₄O₈ requires: C, 63·46; H, 5·76; O, 30·76%).

Tetrahydrobudlein-A (2). A soln of budlein-A (1 g) in MeOH (20 ml)) was hydrogenated for 5 hr with Pd/CaCO₃ (100 mg). The catalyst was filtered off and the soln evaporated to dry-

ness. Purification by preparative TLC afforded 60 mg of tetrahydrobudlein-A; mp 43°; $\lambda_{\rm max}$ 220, 259 nm; ϵ , 7,000 and 10000; $\nu_{\rm max}$ 3450, 1760, 1660 and 1590 cm⁻¹; M⁺ m/e 378.

Octahydrobudlein-A (3). A soln of budlein-A (100 mg) in MeOH (8 ml) was mixed with a cold soln of NaBH₄ (68 mg) in MeOH (2 ml). The reaction mixture was kept at 0° for 1 hr. It was acidified and extracted with CHCl₃. The organic layer was washed, dried and evaporated to dryness. The crude product was recrystallized from AcOEt-hexane; yield 77 mg, mp $164-166^\circ$; $\lceil \alpha \rceil_0^{25} - 168^\circ$ (MeOH); ν_{max} 3430, 1760, 1720 and 1660 cm⁻¹. (Found: C, 62·53; H, 7·62; O, 29·86. $C_{20}H_{30}O_7$ requires; C, 62·82; H, 7·85; O, 29·32%).

Hexahydrobudlein-A acetate (4). A soln of 1b (800 mg) in MeOH (20 ml) containing 300 mg of Pd/C -10% was hydrogenated for 12 hr at 4 kg/cm². The catalyst was filtered off and the soln evaporated to dryness; a mixture of 3 compounds was obtained. Separation by preparative TLC afforded 390 mg of pure hexahydrobudlein acetate; mp 75–78°; $\lambda_{\rm max}$ 258 nm; ε , 12,200. (Found: C, 62-60; H, 7-06; O, 30-40. C₂₂H₃₀O₈ requires: C, 62-55; H, 7-10; O, 30-33%).

Budlein-A epoxide (5). To a soln of 1a (200 mg) in CHCl₃ (20 ml) was added recrystallized m-chloroperbenzoic acid (175 mg) and the soln was allowed to reflux for 48 hr. Work up in the usual manner yielded a mixture of several compounds. Separation by preparative TLC afforded 70 mg of 5 mp 66-69°; M^+ m/e 390.

Budlein-B diacetate (6b). A soln of 6a (52 mg) in Ac₂O (0.5 ml) and C₅H₅N (0.5 ml) was 18 hr at room temp. Work up in the usual manner afforded 50 mg of budlein-B diacetate, mp 149-151°, identical in all respects with zexbrevin D. (Found: C, 65.38; H, 6.82; O, 27.47. C₁₉H₂₄O₆ requires: C, 65.50; H, 6.90; O, 27.60%).

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